



Effect of Dietary *Persea americana* on the Organosomatic Indices, Diseases Resistance and Liver Histopathology of *Clarias gariepinus* Exposed to *Klebsiella pneumoniae*

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Authors' contributions

This work was carried out in collaboration between both authors. Both authors read and approved the final manuscript.

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ABSTRACT

The experiment was carried out to evaluate the prophylactic efficiency of dietary powdered *Persea americana* leaves on the organosomatic indices, disease resistance, and liver histopathology of *Clarias gariepinus* exposed to *Klebsiella pneumoniae*. Five (5) different isonitrogenous diets with varying percentages of *P. americana* powdered leaves inclusion were formulated as follows: Do(0%); D1(3%); D3 (6%); D3 (9%) and D4 (12%). One hundred and fifty (150) *C. gariepinus* were distributed in five groups in triplicates of ten (10), and fed diets Do-D4 accordingly. After eight (8) weeks of feeding, they were injected intraperitoneally with *klebsiella pneumoniae* at days 1, 7, 14 and 21. After twenty eight (28) days post-infection period, three fish from each of the triplicate were sacrificed for evaluation of the Hepatosomatic index (HSI); Cistosomatic index (CSI), and Splenosomatic index (SSI), and the liver was taken to the laboratory for histopathological analysis. The survival rate was calculated in each of the groups and the disease resistance was determined. At the end of the experiment, serious ulcerations were observed on the fish fed Do and infected with *K. pneumonia*, while the fish fed D1 – D4 were ulcer-free. The result reveals that the SSI and CSI were similar in all the groups (Do-D4), but the HSI was higher ($P>0.05$) in the group fed Do (control) compared to the group fed D1-D4. The survival rate and the diseases were

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Lower (<0.5) in the group Do compared to the group fed D1-D4, though it was not dose dependant. The result of the histopathology shows that the liver of the fish fed Do has bloodstain in the portal vein and sinusoid, while the liver of the fish fed D1-D4 have no bloodstain. The experiment reveals that *Persea americana* powdered leaves as applied is an anti-liver inflammatory herb, disease resistant and anti-bacterial.

Keywords: Aquaculture; histology; organosomatic Indices; disease and klensilla pneumonia.

1. INTRODUCTION

Aquaculture is the rearing of aquatic organisms in conducive and controlled environments or enclosures. It is one of the fast-growing food sectors in the world [1]. The demand for fish and fish products has been on the high side due to population increase, and this cannot be met by the supply from natural water bodies. Environmental degradation and population increase have affected aquaculture in developing countries [2], but agriculture and science have been developing ways to enhance productivity in aquaculture to meet up the high population demand and reduce the effects of environmental degradation [3]. Nutrients contained in fish and fish products could go a long way to solve the world's malnutrition problems [4], and more than half a billion people all over the world are relying on aquaculture and fisheries for a living [5].

One of the problems associated with aquaculture practice is the presence of microorganisms. And this is a result of so many factors, including maladministration of feed [6]. The presence of micro-organisms makes fish susceptible to diseases, which cause death and retard growth [7], and most of these diseases are caused by bacteria, viruses, fungi, parasites, etc. [8]. There is a serious need to improve disease resistance in aquaculture to enhance feed acceptability and growth [9,10].

Synthetic drugs and chemicals have been used in aquaculture for the prevention and eradication of diseases, but their uses are associated with disadvantages such as polluting the environment, depositing on fish flesh, drug resistance, etc [10]. The use of herb and herbal products is fast replacing the use of chemicals, because it does not pollute the environment, deposit on fish flesh, and are not immunospecific [10].

The presence of disease or pathogen causing disease can be assessed in fish in so many ways: Haematological analysis [11,12]; Organosomatic indices [13,14], Biochemical

parameters [15,16] Histopathological analysis [17] among others.

Several herbs and herbal extracts have shown their potency as antibacterial in aquaculture, they include *Caricapapay* aqueous root extracts [18], *Persea americana* aqueous leaves extracts [19], *Azardirachta/curcuma lionga* aqueous extracts [20], etc.

The *Persea americana* leaves extracts have been reported to contain medicinal phytochemicals [21, 22]. This research work tends to assess the efficacy of *P. americana* powdered leaves as in *Clarias gariepinus* infected with *Klebsiella pneumonia*.

2. MATERIALS AND METHODS

2.1 Experimental Fish

The experimental fish was purchased in Idi-Onyana farms along Abua Ahoada road in Abua/Odual Local Government Area of Rivers State.

2.2 Experimental Diets

Persea americana leaves were harvested within Port Harcourt, Rivers State, Nigeria. They were air-dried to constant weight, grounded to powder, sieved, and stored. The sieved powdered leaves were added to 38.35% Cp formulated diets Do, at 3%(D1), 6% (D2), 9% (D3), and 12% (D4) respectively following the method of [23].

2.3 Experimental Procedure

150 catfish (117.80 ± 0.11 g and 25.88 ± 0.14 cm) were stocked in the experimental tanks in triplicate at 10 fish per tank. Feeding commenced 24 (twenty-four) hours after stocking, and the fish were fed with the experimented diets (Do – D4) accordingly at 5% body weight per day, two (2) times daily. After 8 weeks of feeding, the fish were infected

intraperitoneally with *klebsiella pneumoniae* at days 1, 7, 14 and 21 with 1.5ml of 1.9×10^5 cfu/ml of overnight grown *K. pneumoniae*. After 28 days post-infection, the liver, spleen, heart of three fish in all the triplicates were harvested for organosomatic indices, the liver was also analyzed for histopathology, and the disease resistance ability and survival rates were determined in each group

2.4 Determination of Organosomatic Indices

This was determined using the formula

$$\frac{\text{Weight of Organ}}{\text{Weight of fish}} \times 100 \quad [15]$$

2.5 Histopathological Analysis

The fish liver was taken to the laboratory in sample bottles containing 10% formalin solution. The samples (liver) were manually processed and trimmed using a rotary microtome (LEICA RM 2125 RTS), manufactured by LEKA Brosysteo, Buffalo Grove, U.S.A. Tissues were dewaxed, stained in hematoxylin and eosin for a display of tissue architecture. Stained slides were examined under light microscope at x 10 magnification.

2.6 Anti-pathogenic Ability (Disease Resistance)

This was determined as relative percentage survival, using the formular:

$$\text{RSP} = 1 - \frac{\% \text{ Mortality in treated}}{\% \text{ Mortality in control}} \times 100 \quad [24]$$

Where RSP = Relative Survival Percentage

2.7 Data Analysis

The collected data were analyzed using SPSS statistics software 17.0 windows. A one-way analysis of variance (ANOVA) was employed to reveal a significant difference between control and treated groups. Tukey's multiple comparison test was applied to separate treatments with significant differences [25].

3. RESULTS

The result of the Organosomatic indices is shown in Table 1, there was no significant difference in

the spleenosomatic and cardiosomatic indices in the experimental fish fed Do – D4. The hepatosomatic index was significantly higher in fish fed Do, but significantly the same in fish fed D1 – D4 diets. The result for the disease resistance fluctuated across the diets, but it was significantly lower in the fish fed Do (control) followed by the fish fed D1 and significantly the same in the fish fed D2 – D4 (Table 2).

Plates 1 (a-f) shows the liver histopathology of the experimental fish infected with *K. pneumoniae*. **1a** is the liver of fish fed Do without infection, it has normal portal vein (P) and there are patches of liver vacuolations; **1b** is the liver of fish fed Do and infected with *K. pneumoniae*, it has blood- stained portal vein with micro liver vacuoles and sinusoid; **1c** is the liver of fish fed D1 and infected with *K. pneumoniae*, it has central vein (CV) with no liver vacuoles and sinusoid; **1d** is the liver of fish fed with D2 and infected with the *K. pneumoniae*, it has micro liver vacuoles and sinusoid; **1e** is the liver of fish fed D3 and infected with *K. pneumoniae*, it has a portal vein, micro vacuoles, and sinusoid; while **1f** is the liver of fish fed D4, it has patches of liver vacuolations. Fig. 1 shows a picture of fish fed Do and infected with *K. pneumoniae* and Fig. 2 shows a picture of fish fed supplemented diets and injected with *K. pneumoniae*.

4. DISCUSSION

4.1 Behaviour Observation

There were no ulcerations on the body of the fish fed D1 – D4 after the infection with *K. pneumoniae*, but there was serious ulceration in the fish fed Do and exposed to *K. pneumoniae*. Similar result was obtained when *C. gariepinus* was challenged with *P. aeruginosa* and exposed to *Carica Papaya* aqueous root extracts [11]. [26] also reported similar result when *punica* peel and oxytetracyclia were administered on *A. hydrophila* infected *C. gariepinus*. This could be as a result of the phytochemicals present in the *P. americana* powdered leaves that are antibacterial [27-29] also reported that phenolic compounds (as contained in *P. americana* in this work) seriously inhibit microbial activities. The result obtained in this work could also be as a result of improved thrombocytes formation in fish fed D1 – D4, which is believed to prevent injuries in infected fish [30;13].

4.2 Organosomatic Indices

Organosomatic indices such as HSI, SSI, and CSI are used to evaluate the health status of fish and other organisms [31,14,32]. Some of the factors that affect the organ somatic indices in fish include water quality, feed type, presence of pathogen/disease, etc. [14,32,13]. The result of the organ somatic indices after the period of infection in this work reveals that there was no significant difference in the CSI and SSI in the experimental fishes but the HSI value was higher

($P>0.05$) in the fish fed Do compare to the fish fed D1 – D4. This result is in agreement with the reports of [33] who reported an increase in HSI when *Rattus rattus* was infected with *cysticercus fasciolaris* and [13] when *C. gariepinus* fed dietary mango back were experimentally infected the *P. aeruginosa*. [15] also reported an increase in the size of the liver, spleen, and intraperitoneal fat in the control, when *C. gariepinus* was challenged with *P. aeruginosa* and exposed to *P. americana* aqueous extracts.

Table 1. Organosomatic indices of some organs of *Clarias gariepinus* fed with Avocado pear leaf Supplemented diets and infected with *Klebsiella pneumonia* (Mean ±SD)

Parameters	Diets				
	D0	D1	D2	D3	D4
HSI	2.00±0.25 ^a	1.71±0.09 ^b	1.06±0.21 ^b	1.19±0.02 ^b	1.20±0.08 ^b
CIS	0.08±0.02 ^a	0.15±0.00 ^a	0.78±0.01 ^a	0.10±0.02 ^a	0.06±0.05 ^a
SSI	0.06±0.04 ^a	0.08±0.04 ^a	0.06±0.02 ^a	0.07±0.01 ^a	0.03±0.02 ^a

Means within the same roll with different superscripts are significantly different ($p<0.05$)
 HSI: Hepetasomatic Index; CSI: Cardiosomatic Index and SSI: Spleenosomatic Index

Table 2. Percentage survival and relative survival percentage (Disease Resistance) of *Clarias gariepinus* fed with avocado pear leaf supplemented diets and infected with *Klebsiella pneumonia* (Mean ±SD)

Parameters	Diets				
	D0	D1	D2	D3	D4
% Surv	66.66±5.77 ^c	90.00±10.00 ^b	96.66±5.77 ^a	96.66±5.77 ^a	93.33±11.54 ^a
RSP	0.00±0.00 ^d	69.33±33.85 ^c	91.66±14.43 ^a	91.66±14.43 ^a	77.66±38.68 ^b

Means within the same roll with different superscripts are significantly different ($p<0.05$) % Surv: Percentage Survival; RSP: Relative Survival Percentage



Fig. 1. Fish Fed Do and Infected with *K. pneumonia*

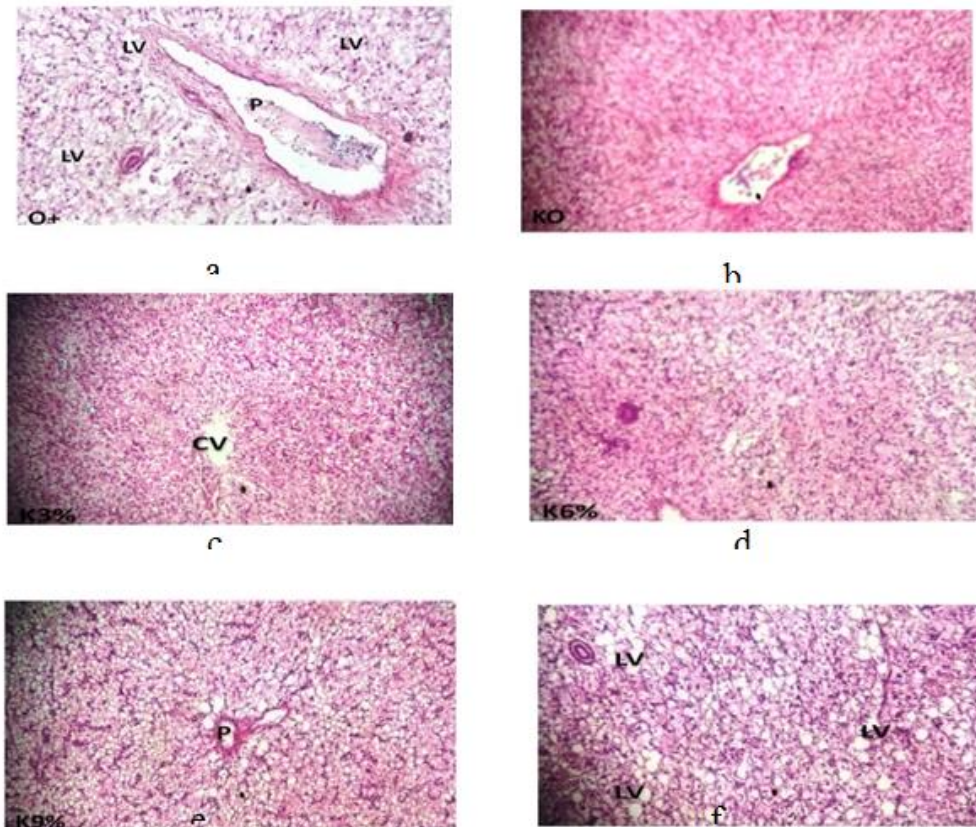


Plate 1. Pathology of liver of *C.gariepinus* fed the experimented diets and exposed to *Klebsiella Pneumonia*

- a: liver of fish fed (Do) 0% leaf inclusion diet without infection
- b: liver of fish fed (Do) 0% leaf inclusion diet exposed to *K.pneumoniae*
- c: liver of fish fed (D1) 3% leaf inclusion diet exposed to *K.pneumoniae*
- d: liver of fish fed (D2) 6% leaf inclusion diet exposed to *K.pneumoniae*
- e: liver of fish fed (D3) 9% leaf inclusion diet exposed to *K.pneumoniae*
- f: liver of fish fed (D4) 12% leaf inclusion diet exposed to *K.pneumoniae*



Fig. 2. Fish Fed *P. americana* supplemented diet and infected with *K. pneumonia*

The increased HSI in the infected fish fed Do (control) and infected with *K. pneumoniae* could be as a result of glycogen depletion [34] arising from the restlessness and rapid opercula movement observed in the fish fed Do when they were injected with the *K. pneumoniae* and depletion of glycogen leads to liver inflammation [35]. Lack of glycogen also leads to the metabolic formation of fat in the liver [36] and this could lead to liver weight increase. The lower HSI in the fish fed D1-D4 compared to fish fed Do could be as a result of the antibacterial activities in the experimental herb [37;18]. It could also be a result of the anti-inflammatory effects of flavonoids and Saponins present in the *P. americana* powdered leaves [38; 15].

4.3 Liver Histopathology

There was a high presence of liver vacuoles in the liver of fish fed 0% leaf inclusion diet without exposure to *K. pneumoniae* (positive control), but the liver of the fish fed 0% leaf inclusion diet and exposed to *K. pneumoniae* (negative control) had no vacuoles, but minor necrosis. This result is similar to the result of [38] when *Lactobacillus acidophilus* was used as a biocontrol agent against *Clarias gariepinus* juveniles infected with pathogenic bacteria (*Aeromonas hydrophila*). The liver of the fish fed D1 – D4 and exposed to *K. pneumoniae* had little presence of liver vacuolation, with more in the fish fed D4. The high presence of liver vacuolation in the positive control could be as a result of the presence of high glycogen due to energy intake and use, as a ($P<0.05$) in fish fed Do compare to the fish fed D1-D4, though it was not dose dependent. The result is similar to the reports of [13] who reported the effect of mango bark extract on *C. gariepinus* infected with *P. aeruginosa*; [46] who reported the effect of neem leaf powder on common carp infected with *A. hydrophila*. [47] also reported high disease resistance when *P. aeruginosa* infected *C. gariepinus* was exposed to *Carica papaya* aqueous extracts and attributed the results to the presence of phytochemicals such as phenols. The high disease resistance in the fish fed D1-D4 could be as a result of the phytochemicals such as flavonoids and other alkaloids present in the *P. americana* leaves that are bactericidal and bacteriostatic [18; 21].

4.4 Disease Resistance

The disease resistance ability which was calculated as relative percentage survival (RSP),

and the percentage survival were low ($P<0.05$) in fish fed Do compare to the fish fed D1-D4, and it increased as the percentage inclusion of *P. americana* leaves increased in the diets. The result is similar to the reports of [13] who reported the effect of mango bark extract on *C. gariepinus* infected with *P. aeruginosa*; [46] who reported the effect of neem leaf powder on common carp infected with *A. hydrophila*. [47] also reported high disease resistance when *P. aeruginosa* infected *C. gariepinus* was exposed to *Carica papaya* aqueous extracts and attributed the results to the presence of phytochemicals such as phenols. The high disease resistance in the fish fed D1-D4 could be as a result of the phytochemicals such as flavonoids and other alkaloids present in the *P. americana* leaves that are bactericidal or bacteriostatic [18; 21].

5. CONCLUSION

The result of this research work shows that powdered *P. americana* is a good prophylactic agent against ulceration and other skin diseases in *Clarias gariepinus* that may arise from the attack of *K. pneumoniae*. Despite being a good antibacterial as seen as the result of the disease resistance in this work, *P. americana* powdered leaves have also proven to be a good anti-inflammatory and anti-necrosis agent in *C. gariepinus* liver in the presence of pathogens such as *K. pneumoniae*. *P. americana* powdered leaves should also be tested for their efficacy against other fish pathogens, to enhance their usage in aquaculture. To this end, we advised that *P. americana* trees should be planted in our farms for the availability of the leaves when needed.

DISCLAIMER

The products used for this research are commonly and predominantly use products in our area of research and country. There is absolutely no conflict of interest between the authors and producers of the products because we do not intend to use these products as an avenue for any litigation but for the advancement of knowledge. Also, the research was not funded by the producing company rather it was funded by personal efforts of the authors.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Food and Agriculture Organization (FAO). The state of food insecurity in the World 2013. The multiple dimensions of food security. Rome; 2013.
2. Otene BB, Ukwe OIK. Evaluation of Heavy Metal Accumulation in Water and Sediment from Elechi Creek, Port Harcourt, Nigeria International Journal of Geography and Environmental Management. 2018;4(1):1-9.
3. Akinrotimi OA, Gabriel UU, Edun OM. The efficacy of clove seed extracts as an anaesthetic agent and its effect of haematological parameters of African Catfish (*Clarias gariepinus*). International journal of Aquaculture and fish Science. 2015;1:012 – 047.
4. Fasakin EA. Fish as food yesterday, today and forever. Inaugural lecture series 48. The Federal University of Technology, Akure; 2007.
5. Future Directions International. Strategies Analysis paper: Fish for the future and food security. Independent strategic analysis of Autalia's Global Interests; 2013.
6. Ukwe IOK, Jamabo NA. Effects of Dietary Mango Bark (*Mangifera indica*) Extract on *Clarias gariepinus* (Burchell, 1822) Infected with *Pseudomonas aeruginosa*. World Journal of Fish and Marine Sciences. 2020;12(3):74-80.
7. Edward JN. Fish Diseases: Diagnosis and Treatment. Second edition, Wiley Blackwell; 2010.
8. Sharma M, Shrivastav AB, Sahni YP, Pandey G. Overviews of the Treatment and Control of common fish diseases. International Research Journal of Pharmacy. 2012;3(7):123-127.
9. EL-Haroun ER, Ma A, Goda S, Kabir CMA. Effects of dietary probiotics Biogens supplementation as growth promoters on growth performance and feed utilization of Nile tilapia, *Oreochromis niloticus*. Aquatic Research. 2008;37:1473 – 1480.
10. Ukwe IOK, Gabriel UU. Herbs and Herbal supplements: Key to a productive, Healthy and Eco-friendly aquaculture. Delta Agriculturist. 2019;11(1/1):55 – 67.
11. Ukwe OIK, Abbey IS, Akinrotimi OA. Behavioural and Haematological Changes in *Pseudomonas aeruginosa* Infected *Clarias gariepinus* Exposed to *Carica papaya* Root Extracts. Hematology Asian Research Journal. 2021;5(1):20-33.
12. Alsaid M, Abuseliana AF, Deud HH, Mustapha NM, Bejo SK, Abdelhadi VM, Handan RH. Haematological, Biochemical and Clinical signs changes following Experimental Infection of *Streptococcus agalactiae* in Red Hybrid Tilapia (*Oreochromis sp.*). Aquaculture Indonesiana. 2014;15(2):86-93.
13. Ukwe IOK, Jamabo NA. Effects of Dietary Mango Bark (*Mangifera indica*) Extract on *Clarias gariepinus* (Burchell, 1822) Infected with *Pseudomonas aeruginosa*. World Journal of Fish and Marine Sciences. 2020;12(3):74-80.
14. Adeniran A, Adeyemo OK, Emikpe BO, Alarape SA. Organosomatic Indices, Haematological and Histological Assessment as Biomarkers of Health Status in Feal and Cultured *Clarias gariepinus* African Journal of Biomediclcs Research. 2017;20:189-194.
15. Ukwe IOK, Etire DI. The Effects of *Persea americana* leaves on the Enzymes and Organosomatic indices of *Pseudomonas aeruginosa* Infected *Clarias gariepinus* Journal of Medical Care Research and Review. 2021;4(7):1-29.
16. Gabriel NN, Quiang J, Ma XY, He J, Xu P, Liu K. Dietary Aloe vera improve plasma liquid profile, antioxidant, and hepatoprotective enzyme activities in GIFT-tilapia (*Oreochromis niloticus*) after *Streptococcus iniae* challenge. Fish physiology and Biochemistry. 2015;4:1321 – 1332.
17. El-Barbary MJ. Some clinical, microbiological and molecular characteristics of *Aeromonas hydrophila* isolated from various naturally infected fishes Aquaculture International. 2010;18: 943-954.
18. Ukwe IOK, Vopnu FB. Diseases Resistance and Enzymatic Changes in *Pseudomonas aeruginosa* Infected *Clarias gariepinus* treated with *Carica Papaya* root extracts. Journal of Medical Care Research and Review. 2021;4(7):1-26.
19. Ukwe IOK, Irondi P. Effects of *Persea americana* leaves extracts on antioxidant activities and diseases resistance of *Pseudomonas aeruginosa* infected *Clarias gariepinus*. European Journal of Biomedical and Pharmaceutical Sciences. 2021;8(9):67-78.
20. Harikrishnan R, Balasundaram C, Heo MS. Potential use of probiotic and triherbal extract – enriched diets to control

- Aeromonas hydrophila infection in carp. Diseases of Aquatic Organisms. 2010;92:41-49.
21. Ogundare AO, Oladejo BO. Antibacterial Activities of the leaf and Bark Extract of *Persea Americana*. American Journal of Ethnomedicine. 2014;1(1):64-71.
 22. Oyeyemi AO, Oyeyemi RB. Effect of Aqueous of the leaves and seeds of Avocado pear (*Persea Americana*) on some Marker Enzymes and Cholesterol in Albino Rat Tissues. Journal of Environmental Science, Toxicology and food Technology. 2015;9(3):15-18.
 23. Lee DH, Ra CS, Song YH, Sung KI, Kim JD. Effects of dietary garlic extract on growth, feed utilization and whole body composition of juvenile starlet sturgeon (*Acipenser ruthenus*). Asian Australian Journal of Animal Sciences. 2012;25(4):577-583.
 24. Harikrishnan R, Kim JS, Balusundaram C, Heo MS. Protection of vibrio herveyi infection through dietary administration of *Pueraria thunbergiana* in kelp grouper, *Epinephelus bruneus*. Aquaculture. 2012;324:27-32.
 25. Wahua TAT. Applied statistics for scientific studies. Africa links books. Aba, Nigeria. 1999;365.
 26. Reda MR, Galal AAA, Alam RTM. Comparism of *Punica Peel* and Oxytetracy on *Aeromonas hydrophila* challenged *Clarias gariepius*. Global Journal of Fisheries and Aquatic Research. 2012;5(5):01-15.
 27. Noorul H, Nesar A, Zafar K, Khalid M, Zeeshan A, Vartika S. Health benefits and pharmacology of *Persea Americana* mill (*Avocado*). International Journal of Research in Pharmacology and Pharma cotherapeutics. 2016;5(2):132-141.
 28. Sales CH, Souza PR, Peghini BC, Da Silver JS, Cardoso CR. An overview of the modulatory effects of Oleic acid in health and diseases. Mini-reviews in Medicinal Chemistry. 2013;13:1-11.
 29. Vasconcelos LC, Sampaio FC, Sampaio MC, Pereira Mdo S, Higino JS, Peixoto MH. Minimum inhibitory concentration of adherence of *Punica grantum* Linn (pomegranate) gel against *S. Mitans* S, *Mitis* and *C. albicans*. Brazilian Dental Journal. 2006;17:223 – 227.
 30. Etim L, Ekanem SB, Utim A. Haemetological profile of two species of catfish *chrysthus nigrodigitatus* and *chrysthus furcatus* from the great kwa River, Nigeria. Global Journal of Pure and Applied Science. 1999;5(1):1-8.
 31. Gabriel UU, Obomanu FG, Edori OS. Haematology, Plasma enzymes and Organ indices of *Clarias gariepinus* after intracellular injection with aqueous leaves extracts of *Lepidagathis alopecurotides* African Journal of Biochemical Research. 2009;3:312-316.
 32. Kareem ZH, Abdelhadi YM, Christianus A, Karim M, Romano M. Effects of some dietary crude plant extracts on the growth and gonadal maturity of Nile tilapia (*Oreochroius niloticus*) and their resistance to streptococcus agalactiae infection. Fish Physiology and Biochemistry. 2016;42:757-769.
 33. Gupta N, Gupta DK, Sharma PK. Condition factor and organosomatic indices of Parasitized *Rattus rattus* as indications of host health. Journal of Parasitic diseases. 2016;61:21-25.
 34. Quentel C, Obach A. The cellular composition of the blood and hematopoietic organs of turbot, *Scophthalmus maximus*. Journal of Fish Biology. 1992;415:709-716.
 35. Bandsman RH, Prinsen BH, Van DV, Rake JP, Boer T, Smith GO, Reingngoud DJ, Kuipers F. Increase de novo lipogenesis and delayed conversion of large VLDL into ointermediate density lipoprotein particles contribute to hyperlipidenua in glurogen storage disease type. La pediatric Research. 2008;63:702-707.
 36. Irima JM, Meyer CM, Segvich DM, Surendran S, Depali – Roach AA, Moral N, Roach PJ. Lack of liver glycogen causes hepatic insulin resistance and steatosis in mice. Journal of Biological Chemistry. 2018;292 (25):10455–10464.
 37. Bowser PR, Wooster GA, Chen CY, Mo RS. Polymicrobial infection of hybrid striped bass (*Morone chrysops* x *Morone saxatilis*) with three bacterial pathogens: A case report. Journal of Fish Diseases. 2004;27:123 – 127.
 38. Wolf CJ, Wolfe MJ. A Brief Overview of Nonneoplastic Hepatic toxicity in fish. Toxicologic Pathology. 2005;33:75-85.
 39. Boorman GA, Batts S, Bunton TE, Fournie JW, Harshbarger JC, Hawkins WE, Hinton DE, Diagonistic criteria for degenerative, inflammatory, Proliferative non-neoplastic and neoplastic liver lesions in medaka (*Oryziaslatipes*) consensus of a National Toxicology program pathology Working

- Group. Toxicology Pathology. 1997;25:202-210.
40. Adeyemi JA. Oxidative stress and antioxidant enzymes activities in the African catfish. *Clarias gariepinus*, experimentally challenged with *Eshenchina coliarod vibro fischeri*. *Fish Physiology and Biochemistry*. 2014;40:347-354.
 41. Penrith MI, Bastianello SS, Penrith MJ. Hepatic lipidosis and fatty infiltration of organs in captive African Stonefish, *Synanceja verrucosa* Bloch and Schneider. *Journal of Fish Diseases*. 1994;17:171-176.
 42. Ojewole JA, Amabeoku GJ. Anticonvulsant effect of *Persea americana* mill, Lauraceae (Avocado) leaf aqueous extract in mice. *Phytotherapy Researcher*. 2006;20(8):696 – 700.
 43. Brai B, Adisa AR, Odetola AA. Hepatoprotective properties of Aqueous leaf Extract of *Persea Americana*, Mil (*Laura ceac*) “Avocado” Against Cel <Sub>4,/Sub> Iduced Damage in Rats. *African Journal of Traditional Complementary and Alternative Medicine*. 2014;11(2):236 – 244.
 44. Verma RK, Kumari M, Singh G. Ameliorating effect of neem (*Azadiracha indica*) leaf powder on pathology of *Aeromonas hydrophila* infection in common carp (*Cyprinus carpio* L.) *Annals of Biology*. 2013;29:418 – 428.
 45. Sivaram V, Babu MM, Citarasu T, Immanuel G, Murugadass S, Mariam MP. Growth and Immune response of juvenile greasy groupers (*Epinephelus tavivina*) fed with herbal antibacterial active principle supplemented diets against *Vibro harveyi* infections. *Aquaculture*. 2004;237:9 -20.
 46. Kumala S, Utami H, Sari WK. The effect of Avocado (*P. americana* mill) leaves Extract Towards the Mouse’s Blood Glucose Decrease with the Glucose Tolerance Method. *International Journal of Pharmaceutical Sciences and Research*. 2013;4(2):661 – 665.
 47. Al-Dohail MA, Hashim R, Aliyu-Paiko M. Evaluating the use of *Lactobacillus accidephilus* as a biocontrol agent against common pathogenic bacteria and the effects on the haematology parameters and histology in African catfish *clarias gariepinus* juveniles. *Aquaculture Research*. 2011;42(2):196-209.

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