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Determination of Polyphenols, Reducing Potential and FT-IR Analysis of Green Tea (Camellia sinensis L.) Leaves Extract for Nanoparticles Synthesis

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Authors' contributions

This work was carried out in collaboration among all authors. Author KB performed the methodology, investigated and analysed the data, wrote original draft and edited the manuscript. Author NP wrote and edited the manuscript. Author SG conceptualized the study, reviewed and edited the manuscript. Author PL supervised and conceptualized the study, searched for resources, reviewed and edited the manuscript. All authors read and approved the final manuscript.

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ABSTRACT

Green Tea (*Camellia sinensis* L.) is renowned for its therapeutic properties due to its phenolic compounds. Extracts from plants with high polyphenol content possess antioxidant properties that neutralize free radicals involved in the oxidation of various molecules. This study investigated the

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total phenolic and flavonoid content, 1,1-diphenylpicrylhydrazyl (DPPH) radical scavenging activity (RSA), and ferric reducing antioxidant power (FRAP) of commercially available green tea leaves for potential use in the green synthesis of silica nanoparticles. The antioxidant activity of green tea leaves demonstrates its reducing power, which is crucial for utilizing its extract in nanoparticle synthesis. Gallic acid and quercetin served as standards in the Folin–Ciocalteu and aluminum chloride methods, respectively, to measure total phenolic and flavonoid concentrations. To determine antioxidant content ascorbic acid and gallic acid were standards for the DPPH free radical scavenging and FRAP assays. Fourier Transform Infrared (FT-IR) spectra was performed to evaluate the green tea extract. The results revealed green tea leaves exhibited high phenolic, flavonoid content and significant antioxidant activity. The presence of few notable vibrational bands in FT-IR graph also confirmed the existence of chemical substances that are associated with antioxidant qualities (polyphenols, flavonoids). Indicating that the green tea leaf extract can act as an effective reducing and capping agent in the synthesis of silica nanoparticles.

Keywords: Camellia sinensis L.: green tea extract: phenol: flavonoid: antioxidant activity: FT-IR.

1. INTRODUCTION

Tea plant, (Camellia sinensis) is indigenous to Asia and a member of the Theaceae family. It is cultivated extensively all around the world and used to make the commercialized product green tea [1]. Green tea is an intricate blend of polyphenols which are vital biologically active substances [2]. The leaves, which serve as the primary ingredient in the creation of various varieties of tea, are abundant in polyphenols, they additionally contain a range of commonly known proteins, amino acids, alkaloids, and phytochemicals such as caffeine, vitamins C and E, mineral compounds (such as potassium, fluoride. and aluminum). phenolic acids. condensed tannins, and hydrolyzable tannins, trace elements (such as zinc, magnesium, and folic acid), lipids, pigments, and aromatic compounds [3]. Of these, the two main types of polyphenols present in green tea are flavanols and flavonols. Flavonoids and their glycosides, chlorogenic acid, gallic acid, coumarylquinic acid, and theogallin are some of the additional polyphenols that are found. Gallocatechins are catechins characterized by having three hydroxyl groups at position 3 of the ring, whereas the replacement of gallic acid at the same position defines catechin gallates [3]. Favonols, also known as catechins, consist of hydroxyl groups and aromatic rings, which grant them strong antioxidant properties [4]. Flavonoids are polyphenols characterized by a 15-carbon skeleton, with flavonols as one of their subgroups [5]. Together, they account for approximately 16-30% of the weight of the fresh tea leaf when it is dried [6,7].

The flavan-3-ols, specifically epigallocatechin (EGC), epigallocatechin-3-gallate (EGCG),

epicatechin (EC), and epicatechin-3-gallate (ECG), have a significant role in the antioxidant properties of green tea (GT) [8]. The catechins, which are colorless, astringent, and watersoluble, are the major species that are rapidly oxidizable. Antioxidants are known as steady substances that furnish electrons to free radicals, stabilizing them and preventing their reactivity with other molecules [9]. "Free radical" refers to molecules that have no electron pair in their atomic orbital, making them very reactive and unstable [10]. Oxidative stress results from an unequal ratio of antioxidants to free radicals within cells or tissues, leading to damage to lipids, proteins, and nucleic acids [11]. To achieve stability, free radicals can either receive or donate electrons (oxidants or reductions) to other molecules, transforming the molecule into a free radicle [12].

Numerous assays are available to measure antioxidant activity, total flavonoid concentration, and total phenolic content [13]. Several studies have investigated the relationship between total flavonoids, phenolics, and antioxidant activity, suggesting a potential linear correlation between these variables [14]. Different methods are employed to assess the antioxidant capacity and overall reducing ability of organisms like plants and algae [15,16]. These techniques belong to the categories of electron transfer and hydrogen atom-based techniques. Tests based on electron transfer include DPPH, Folin-Ciocalteu, and FRAP [17,18]. In these redox reactions, antioxidants in the sample donate electrons to the metal ion in the Folin-Ciocalteu reagent or oxidants like the DPPH radical. Therefore, the complete reducing ability of a sample can be ascertained bγ evaluating changes absorbance at a specific wavelength against a

standard curve [19]. The phenolic compounds' scavenging ability may come from the active hydrogen-donating capacity of the hydroxyl groups [20]. Additionally, the radical scavenging activity is more significantly influenced by high molecular weight, the number and proximity of aromatic rings, and the type of substitution of hydroxyl groups, as opposed to certain functional groups [21].

are several wavs to svnthesize nanoparticles, including chemical, biological, and physical techniques. Reducing and capping agents are essential for the synthesis of nanoparticles [22]. High radiation, highly dangerous, concentrated, and extremely chemicals poisonous are utilized as environmental-harming reducing and stabilizing agents in chemical and physical processes [23]. The green synthesis method, which uses plant extracts to create nanoparticles, is a very costeffective, ecologically friendly, and sustainable procedure. Because green synthesis is a onestep process, the nanoparticles produced have adequate size, greater stability, and a diversity of different natures [24]. During the synthesis from corresponding aqueous salts, a redox reaction occurs [22]. Reducing compounds found in a variety of organism extracts transmit electrons to metal ions, resulting in the formation of nanoparticles [25]. Plants with higher levels of total reducing compounds are expected to yield increased concentrations of nanoparticles [26].

2. MATERIALS AND METHODS

The following reagents were used: Methanol carbonate Sodium anhydrous (HiMedia). (HiMedia), Gallic acid monohydrate (HiMedia). Sodium nitrite (HiMedia), Sodium Hydroxide (HiMedia), L-Ascorbic acid (HiMedia), Hydrochloric acid (HiMedia), Ferric chloride hexahydrate (HiMedia), Sodium acetate anhydrous (HiMedia), 2,4,6-Tripyridyl-S-Triazine (Central Drug House), 2,2-Diphenyl-1-Picrylhydrazyl (Sigma-Aldrich), Folin Ciocalteus Phenol (Sisco Research (Sisco Laboratories), Acetic Acid Glacial Research Laboratories), Aluminium Chloride Hexahydrate (Sisco Research Laboratories), (Sisco Quercetin Dihydrate Research Laboratories). All reagents used were of Analytical grade. The plant material used was a commercial brand of green tea (GT), obtained from the market and used as the sample for each test. To enhance extraction efficiency, the dried GT leaves were crushed using an

autoclaved mortar and pestle. For the preparation of methanolic extract 2 gram of powdered green tea leaves was added in 100 ml methanol and kept at room temperature in the dark for 24 hr. The extract was filtered through Whatman No.1 filter paper, and stored in refrigerator for further use.

2.1 Total Phenolic Content (TPC)

The green synthesis of nanoparticles is facilitated by phenolic compounds due to their high antioxidant activity, which makes them effective reducing agents. The total phenolic content (TPC) of the sample was assessed [27]. 0.5 mL of methanolic extract was combined with 2.5 mL of 10% Folin-Ciocalteu reagent and 2.5 mL of 7.5% Na₂CO₃ solution. The mixture was then incubated at 45°C for 45 minutes. After incubation, the absorbance of samples at 765 nm was measured using methanol as the blank. The TPC was determined by extrapolating from a calibration curve created using a gallic acid solution. The estimation of phenolic compounds was done in three sets. The average TPC was expressed in milligrams of gallic acid equivalents (GAE) per gram of dried sample.

2.2 Total Flavonoid Content (TFC)

Flavonoids are essential as reducing and electrostatic agents in the synthesis of nanoparticles from their precursors, facilitating the eco-friendly production of nanoparticles (NPs) [28]. The total flavonoid content (TFC) was measured using the aluminum chloride test method [29]. 0.2 mL of 5% NaNO2 was combined with 2 mL of methanolic extract. After five minutes, 0.2 mL of 2% AICl3 was added to the mixture and allowed to stand for six minutes. The entire volume was then adjusted to 5 mL after adding 2 mL of 1N NaOH and the mixture was incubated at room temperature for 15 minutes. Methanol was used as a blank. The absorbance was measured at 510 nm. Quercetin served as the standard reference for quantifying total flavonoids. The total flavonoid content of each extract is expressed in (mg QE) per g of dry weight.

2.3 2,2-Diphenyl-picrylhydrazyl (DPPH) Radical Scavenging Assay

The free radical scavenging activity was evaluated by measuring the percentage decrease in the initial color concentration [30]. The solution consisted of methanol (3 mL), tea

extract (0.5 mL), and DPPH solution (0.3 mL, 0.5 mM) in methanol. The mixture was incubated in the dark at 25°C for 45 minutes. The methanol solution was used as the control blank. The percentage inhibition of DPPH was calculated by measuring the absorbance at 517 nm.

The following formula was used to calculate the DPPH radical scavenging capacity:

% scavenging capacity = [(Abs Control) – (Abs sample)/(Abs Control)]*100

Where Abs Control = Absorbance of the control Abs sample = Absorbance of the sample

This radical scavenging capacity was stated in ascorbic acid equivalents per gram of dry tea (AAE/g), determined by extrapolation from a calibration curve of L-ascorbic.

2.4 Ferric Reducing Antioxidant Power (FRAP) Assay

The antioxidant strength of green tea was estimated using the following method: The FRAP reagent was prepared by mixing 40 mM HCl in a 10:1:1 ratio with 300 mM acetate buffer (pH 3.6), 20 mM ferric chloride hexahydrate, and 10 mM 2,4,6-Tripyridyl-s-triazine (TPTZ). For every 50 μL of green tea sample, 100 μL of FRAP reagent was used. In the same way, ddH2O (50 µL) was used in place of the tea sample to prepare a This mixture was incubated for 10 blank minutes at 25°C i.e. room temperature. Following incubation, absorbance was measured at 593 nm using UV-visible spectrophotometry [31]. The FRAP value is in milligrams of GAE per gram of dried sample. This value is calculated based on the standard curve of gallic acid.

2.5 FT-IR Analysis

20 grams of GT were added in 500 mL of deionized water to create the tea extract. To extract the tea, the mixture was cooked to 80°C in a water bath. The extract was then cooled and filtered using Whatman filter paper. The filtered solution was gathered and stored in a sterile, dry beaker for FT-IR spectroscopy analysis.

3. RESULTS AND DISCUSSION

3.1 Total Phenolic Content (TPC)

The total polyphenol content was determined using the Folin-Ciocalteu method. The assay

results indicated that the dried leaves of GT contained 11.51±0.36 mg GAE/g phenolic content levels expressed as gallic acid equivalent. Similarly, the other findings also confirmed the presence high content of polyphenols in GT leaf extract [32,33]. Research has shown that dried plant materials exhibit higher levels of antioxidant activity antioxidants like polyphenolics compared to fresh plant materials [34]. This is because the drying process can break down cellular components, allowing phenolic compounds to be released from the food matrix more quickly [35,36]. In dried samples, destructive enzymes are inactivated due to low water activity, resulting significant concentrations of phenolic compounds [37]. Polyphenols are the primary compounds responsible for antioxidant activity, consist of one or more hydroxyl groups attached to an aromatic ring. The highest concentration of polyphenols was found in GT due to the presence of flavan-3-ols such as catechin (C), EGCG, ECG, EGC, and EC [38].

3.2 Total Flavonoid Content (TFC)

The total flavonoid content of green tea was measured using the aluminium chloride colorimetric technique. The assay results showed that green tea leaves contained 4.73±0.31 mg QE/g of flavonoids. Our results showed alignment with data obtained other finding confirming the presence of flavonoid in the green tea also plays a significant role in its antioxidant activity [39]. Bioflavonoids, such as catechin and its derivatives, have exceptional antioxidant properties [40]. Catechins are a class of phenolic and flavonoid molecules that are chemically unstable [41]. Catechins easily oxidize in solution, losing hydrogen atoms and forming oxidized quinone products as well as a semiguinone radical intermediate [22,25].

3.3 2,2-Diphenyl-picrylhydrazyl (DPPH) Radical Scavenging Assay

One of the most often used methods for assessing antioxidant activity is the radical scavenging method known as DPPH. In this technique, antioxidant compounds react with the DPPH solution, a free radical, by donating electrons. This reaction neutralizes the free radical, converting it into a non-radical form [26]. The assay results indicated that green tea has a DPPH radical scavenging capacity of 36.45±2.45 mg AAE/g. GT's powerful antioxidant properties are attributed to the three adjacent hydroxyl

(OH) groups on the β -ring of its catechins, such as gallocatechin gallate (GCG), EGCG, and gallocatechin (GC) [42]. These catechins are more efficient at scavenging free radicals compared to those with only two adjacent OH groups, such as epicatechin (EC) and catechin gallate (CG). Green tea has substantially higher levels of EGCG and EGC [43].

3.4 Ferric Reducing Antioxidant Power (FRAP) Assay

The modified ferric ion-reducing antioxidant power (FRAP) method was utilized to assess the total antioxidant power of green tea. The results revealed significant FRAP levels in green tea was 13.86±0.48 mg GAE/g. Our results also supported the previous studies that the FRAP levels were found to be significantly augmented in green tea [42]. FRAP is a degree of compounds' ability to behave as electron donors and green tea contains numerous chemicals capable of transferring electrons [43,44]. The presence of reductones is associated with its power. reducina Reductones function antioxidants by donating a hydrogen atom to break the chain of free radicals [45]. GT contains various polyphenols, with higher levels attributed to flavan-3-ols such as C, EGCG, ECG, EGC and EC. Phenolic compounds exhibit strong reducing power and can donate an electron to convert ferric ion Fe3+ to ferrous ion Fe2+ [46].

3.5 FT-IR Analysis

The FTIR spectra of GT extract revealed a weak band at 769 cm⁻¹ is the result of out of plane vibrations attributed to aromatic C-H bonds [47]. The absorption bands in the position of 1082 cm⁻¹ can be caused by C-OH stretching which belongs to amino acid or alcohol [48]. The band at 1313 cm⁻¹ is caused by the C-N stretch of amide-I in protein [49]. A C-C stretch (aromatic) band was also detected at 1479 cm⁻¹ [50]. The stretching vibration of the C=C (aromatics) and C=O (carbonyl) bonds found in flavonoids, polyphenols and catechins gallic acid, catechin, or L-theanine is attributed to the strong defined band at 1634 cm⁻¹ [50,48]. The source of the 2115 cm⁻¹ vibrations was C-O stretching [46]. At 2925 cm⁻¹, a band associated with the vibration of the C-H stretch or O-H stretch of a carboxylic acid bond was detected [51]. The broad band seen in GT extracts in the 3284 cm⁻¹ region is indicative of the OH stretching of phenolic and flavonoid components [52,53]. The stretching of the hydroxyl group and O-H bond is responsible for the peaks that occur at 3838 cm⁻¹ [54]. Therefore, it can be seen from the FTIR spectrum that the GT sample contains high levels of proteins, carboxylic acid, amino acids and polyphenols. Flavonoids or phenolic compounds may act as reducing agents, and proteins may act as capping or stabilizing agents to prevent agglomeration and give stability.

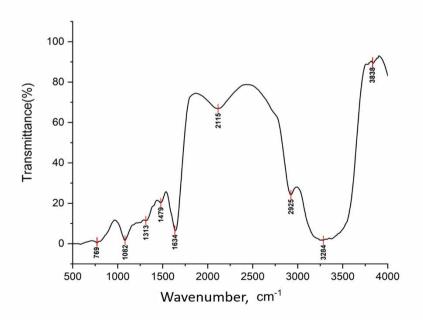


Fig. 1. FT-IR Analysis of Green Tea Extract

4. CONCLUSION

This study showed that the phenolic and flavonoid content of green tea (Camellia sinensis L.) leaf extract demonstrated strong antioxidant activity. The antioxidant properties were FRAP assay, TPH, TFC, DPPH radical scavenging assay. The extract's FTIR spectra displayed bands that corroborated the presence of these chemical components. Thus, it can be concluded that green tea possesses notable antioxidant activity, as well as substantial total phenol and flavonoid content, that will act as reducing leading to the production nanoparticles, indicating the suitability of GT leave extract for the further synthesis of nanoparticles. We believe that this work could stimulate the synthesis of various nanoparticles, that have a wide range of applications.

DISCLAIMER (ARTIFICIAL INTELLIGENCE)

NO Author(s) hereby declare that generative AI technologies such as Large (ChatGPT, Language Models COPILOT, and generators etc) text-to-image have been used during writing or editing of manuscripts.

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COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

- 1. Vargas-Sanchez PK, Pitol DL, de Sousa LG, Beloti MM, Rosa AL, Rossi AC, Siéssere Bombonato-Prado S. rich KF. Green tea extract epigallocatechin gallate impairs alveolar bone loss ovariectomized in experimental periodontal rats with disease. Int. Exp. Pathol. J. 2020;101(6):277-88.
- Luo Q, Zhang JR, Li HB, Wu DT, Geng F, Corke H, Wei XL, Gan RY. Green extraction of antioxidant

- polyphenols from green tea (*Camellia sinensis*). Antioxidants. 2020; 9(9):785.
- Zhang L, Ho CT, Zhou J, Santos JS, 3. Armstrong L, Granato D. Chemistry and biological activities of processed Camellia sinensis teas: Α comprehensive review. Compr. Rev. Food Sci. Food Saf. 2019;18(5):1474-95.
- Wang B, Tu Y, Zhao SP, Hao YH, Liu JX, Liu FH, Xiong BH, Jiang LS. Effect of tea saponins on milk performance, milk fatty acids, and immune function in dairy cow. J. Dairy Sci. 2017 Oct 1;100(10): 8043-52.
- 5. Zhao CN, Tang GY, Cao SY, Xu XY, Gan RY, Liu Q, Mao QQ, Shang A, Li HB. Phenolic profiles and antioxidant activities of 30 tea infusions from green, black, oolong, white, yellow and dark teas. Antioxidants. 2019; 8(7):215.
- 6. Reto M, Figueira ME, Filipe HM, Almeida CM. Chemical composition of green tea (*Camellia sinensis*) infusions commercialized in Portugal. Plant Foods Hum. Nutr. 2007;62:139-44.
- 7. Liang YR, Liu ZS, Xu YR, Hu YL. A study on chemical composition of two special green teas (*Camellia sinensis*). J. Sci. Food Agri. 1990;53(4):541-8.
- 8. Musial C, Kuban-Jankowska A, Gorska-Ponikowska M. Beneficial properties of green tea catechins. Int. J. Mol. Sci. 2020;21(5):1744.
- 9. Nagle DG, Ferreira D, Zhou YD. Epigallocatechin-3-gallate (EGCG): Chemical and biomedical perspectives. Phytochem. 2006;67(17):1849-55.
- Lobo V, Patil A, Phatak A, Chandra N. Free radicals, antioxidants and functional foods: Impact on human health. Phcog. Rev. 2010;4(8):118.
- Caleja C, Barros L, Antonio AL, Oliveira 11. MB, Ferreira IC. A comparative study and synthetic between natural antioxidants: Evaluation of their performance after incorporation into biscuits. Food Chem. 2017; 216: 342-6.
- 12. Phaniendra A, Jestadi DB, Periyasamy L. Free radicals: Properties, sources, targets, and their implication in various diseases. Int. J. Clin. Biochem. 2015;30:11-26.

- Číž M, Čížová H, Denev P, Kratchanova M, Slavov A, Lojek A. Different methods for control and comparison of the antioxidant properties of vegetables. Food Contr. 2010;21(4):518-23.
- Singleton VL, Rossi JA. Colorimetry of total phenolics with phosphomolybdicphosphotungstic acid reagents. Am. J. Enol. Vitic. 1965;16(3):144-58.
- 15. Miller NJ, Rice-Evans CA. Spectrophotometric determination of antioxidant activity. Redox R. 1996;2(3):161-71.
- Schwarz K, Bertelsen G, Nissen LR, 16. Gardner PT, Heinonen MI, Hopia A, Huynh-Ba T, Lambelet P, McPhail D, Skibsted LH, Tijburg L. Investigation of plant extracts for the protection of processed foods against lipid oxidation. Comparison of antioxidant assays based on radical scavenging, biqil oxidation and analysis of the principal antioxidant compounds. Eur. Food Res. Technol. 2001;212:319-28.
- **Everette** 17. JD, QM, Bryant Green Abbey Wangila AM, YA, GW. Walker RB. Thorough study reactivity of various compound classes toward the Folin- Ciocalteu reagent. J. Agri food Chem. 2010;58(14): 8139-44.
- 18. Huang D, Ou B, Prior RL. The chemistry behind antioxidant capacity assays. J. Agri food Chem. 2005;53(6):1841-56.
- Baliyan S, Mukherjee R, Priyadarshini A, Vibhuti A, Gupta A, Pandey RP, Chang CM. Determination of antioxidants by DPPH radical scavenging activity and quantitative phytochemical analysis of Ficus religiosa. Molecules. 2022;27 (4):1326.
- 20. Siddhuraju P, Manian S. The antioxidant activity and free radical-scavenging capacity of dietary phenolic extracts from horse gram (*Macrotyloma uniflorum* (Lam.) Verdc.) seeds. Food Chem. 2007;105(3):950-8.
- Hagerman AE, Riedl KM, Jones GA, Sovik KN, Ritchard NT, Hartzfeld PW, Riechel TL. High molecular weight plant polyphenolics (tannins) as biological antioxidants. J. Agri Food Chem. 1998;46(5):1887-92.

- 22. Thakkar KN, Mhatre SS, Parikh RY. Biological synthesis of metallic nanoparticles. Nanomedicine. 2009;6: 257–262.
- 23. Kuppusamy P. Yusoff MM. Maniam GP, Govindan N. Biosynthesis of nanoparticles metallic using plant derivatives and their new avenues in pharmacological applications updated report. Saudi Pharm J. 2016:(4):473-84.
- 24. Bhardwaj B, Singh P, Kumar A, Kumar S, Budhwar V. Eco-friendly greener synthesis of nanoparticles. Adv Pharm Bull. 2020;10(4):566-576.
- 25. Christensen L, Vivekanandhan S, Misra M, Mohanty AK. Biosynthesis of silver nanoparticles using *Murraya koenigii* (curry leaf): An investigation on the effect of broth concentration in reduction mechanism and particle size. Adv Mat Lett. 2011;2:429–434.
- 26. Song JY, Kim BS. Rapid biological synthesis of silver nanoparticles using plant leaf extracts. Bioprocess Biosyst Eng. 2009;32:79–84.
- 27. Jaradat N, Hussen F, Al Ali A. Preliminary phytochemical screening, quantitative estimation of total flavonoids, total phenols and antioxidant activity of Ephedra alata Decne. J. Mater. Environ. Sci. 2015;6(6):1771-8.
- 28. Bordiwala, R.V. Green Synthesis and applications of metal nanoparticles— A rev. Article. Results Chem. 2023;5:100832.
- 29. Barek ML, Hasmadi M, Zaleha AZ. Fadzelly AM. Effect of different drying phytochemicals methods on and antioxidant properties of unfermented and fermented teas from Sabah Snake Grass (Clinacanthus nutans Lind.) leaves. Int. Food Res. Journal. 2015;22(2):661.
- 30. Moraes-de-Souza RA, Oldoni TL. Regitano-d'Arce Alencar SM. MA, and Antioxidant activity phenolic composition of herbal infusions consumed in Brazil. CYTA-J. Food. 2008;6(1): 41-7.
- 31. Benzie IF, Strain JJ. The ferric reducing ability of plasma (FRAP) as a measure of "antioxidant power": the FRAP assay. Anal. Biochem. 1996;239(1):70-6.
- 32. Ramírez-Aristizabal LS, Ortíz A, Restrepo-Aristizabal MF, Salinas-Villada JF. Comparative study of the antioxidant

- capacity in green tea by extraction at different temperatures of four brands sold in Colombia. Vitae. 2017;24(2):132-45.
- 33. Chakraborty K, Bhattacharjee S, Pal TK, Bhattacharyya S. Evaluation of In vitro antioxidant potential of Tea (Camelia sinensis) leaves obtained from different heights of Darjeeling Hill, West Bengal. J. App. Pharm. Sci. 2015;5(1):063-8.
- Pinela J, Barros L, Dueñas M, Carvalho AM, Santos-Buelga C, Ferreira IC. Antioxidant activity, ascorbic acid, phenolic compounds and sugars of wild and commercial *Tuberaria lignosa* samples: Effects of drying and oral preparation methods. Food Chem. 2012;135(3):1028-35.
- Chang CH, Lin HY, Chang CY, Liu YC. Comparisons on the antioxidant properties of fresh, freeze-dried and hot-air-dried tomatoes. J. Food Eng. 2006;77(3):478-85
- Arslan D, Özcan MM. Evaluation of drying methods with respect to drying kinetics, mineral content and colour characteristics of rosemary leaves. Energy Convers. Manag. 2008;49(5):1258-64.
- 37. Hossain MB, Barry-Ryan C, Martin-Diana AB, Brunton NP. Effect of drying method on the antioxidant capacity of six Lamiaceae herbs. Food Chem. 2010;123(1):85-91.
- 38. Jin L, Li XB, Tian DQ, Fang XP, Yu YM, Zhu HQ, Ge YY, Ma GY, Wang WY, Xiao WF, Li M. Antioxidant properties and color parameters of herbal teas in China. Ind Crops Prod. 2016;87:198-209.
- 39. Fawwaz M, Muflihunna A, Pratama M, Rahmawati R, Razak R, Baits M. Total phenolic and flavonoid compound of crude and purified extract of green tea leaves (*Camellia sinensis*) from Makassar-Indonesia. J. Fitofarmaka Indones. 2022;9(3):19-24.
- 40. Yashin Α, Yashin Y, Nemzer Determination of antioxidant activity in tea extracts. and their total antioxidant content. Am. J. Biomed. Sci. 2011;3(4):322-35.
- 41. Higdon JV, and B Frei. Tea catechins and polyphenols: Health effects, metabolism, and antioxidant functions. Crit. Rev. Food Sci. Nutr. 2003;43:89-143.
- 42. Singh K, Srichairatanakool S, Chewonarin T, Brennan CS, Brennan MA, Klangpetch

- W, Utama-Ang N. Manipulation of the phenolic quality of assam green tea through thermal regulation and utilization of microwave and ultrasonic extraction techniques. Hortic. 2022;8(4):338.
- 43. Almajano MP, Carbo R, Jiménez JA, Gordon MH. Antioxidant and antimicrobial activities of tea infusions. Food Chem. 2008;108(1):55-63.
- 44. Chan EW, Lim YY, Chew YL. Antioxidant activity of *Camellia sinensis* leaves and tea from a lowland plantation in Malaysia. Food Chem. 2007;102(4):1214-22.
- 45. Duh PD. Antioxidant activity of burdock (*Arctium lappa* Linne): Its scavenging effect on free-radical and active oxygen. J. A. Oil Chem. Soc. 1998;75(4):455-61.
- Gordon MH. The Mechanism of Antioxidant Action *In vitro*. In: Hudson, B.J.F., editors. Food Antioxidants. Elsevier Applied Food Science Series. Springer, Dordrecht; 1990.
- 47. Selvan DA, Mahendiran D, Kumar RS, Rahiman AK. Garlic, green tea and extracts-mediated turmeric green synthesis silver nanoparticles: of Phytochemical, antioxidant and In vitro cvtotoxicity studies. J. Photochem. Photobiol. B. 2018;180:243-52.
- Wirwis A. and Sadowski Z. Green synthesis of silver nanoparticles: Optimizing green tea leaf extraction for enhanced physicochemical properties. ACS omega. 2023;8(33):30532-30549.
- Gottimukkala KS, Harika RP, Zamare D. Green synthesis of iron nanoparticles using green tea leaves extract. J. Nanomed. Biother. Discov. 2017;7(1):151.
- 50. Senthilkumar SR. and T. Sivakumar. Green tea (*Camellia sinensis*) mediated synthesis of zinc oxide (ZnO) nanoparticles and studies on their antimicrobial activities. Int. j. pharm. pharm. Sci. 2014;6:461-465.
- 51. Senthilkumar SR. FT-IR analysis and correlation stud phenolics and total flavonoids of India. Int. Res. J. Biological Sci. 2017;6(3):1-7.
- 52. Dubey SP, Lahtinen M, Sillanpää M. Tansy fruit mediated greener synthesis of silver and gold nanoparticles. Process Biochem. 2010;45(7):1065-71.
- 53. Nandiyanto AB, Oktiani R, Ragadhita R. How to read and interpret FTIR spectroscope of organic material. Indones. J. Sci. Technol. 2019;4(1): 97-118.

54. Luceño-Sánchez JA, Maties G, Gonzalez-Arellano C, Diez-Pascual AM. Synthesis and characterization of graphene oxide derivatives via functionalization reaction with hexamethylene diisocyanate. Nanomater. 2018;8(11):870.

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